

Latitude, elevational climatic zonation and speciation in New World vertebrates

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Many biodiversity hotspots are located in montane regions, especially in the tropics. A possible explanation for this pattern is that the narrow thermal tolerances of tropical species and greater climatic stratification of tropical mountains create more opportunities for climate-associated parapatric or allopatric speciation in the tropics relative to the temperate zone. However, it is unclear whether a general relationship exists among latitude, climatic zonation and the ecology of speciation. Recent taxon-specific studies obtained different results regarding the role of climate in speciation in tropical versus temperate areas. Here, we quantify overlap in the climatic distributions of 93 pairs of sister species of mammals, birds, amphibians and reptiles restricted to either the New World tropics or to the Northern temperate zone. We show that elevational ranges of tropical- and temperate-zone species do not differ from one another, yet the temperature range experienced by species in the temperate zone is greater than for those in the tropics. Moreover, tropical sister species tend to exhibit greater similarity in their climatic distributions than temperate sister species. This pattern suggests that evolutionary conservatism in the thermal niches of tropical taxa, coupled with the greater thermal zonation of tropical mountains, may result in increased opportunities for allopatric isolation, speciation and the accumulation of species in tropical montane regions. Our study exemplifies the power of combining phylogenetic and spatial datasets of global climatic variation to explore evolutionary (rather than purely ecological) explanations for the high biodiversity of tropical montane regions.

Keywords: allopatric speciation; diversification; ecological speciation; latitudinal diversity gradient; niche conservatism

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Electronic supplementary material is available at <http://dx.doi.org/10.1098/rspb.2011.0720> or via <http://rspb.royalsocietypublishing.org>.

1. INTRODUCTION

Montane regions are hotspots of species richness and endemism for many taxa, especially in the tropics [1,2]. Tropical mountains are also often outliers in macroecological studies that correlate species richness with present-day climate, suggesting that montane diversity and endemism might be attributed to historical or evolutionary factors in

addition to contemporary ecological processes [3–5]. Yet we know surprisingly little about the processes that lead to increases or decreases in species richness (i.e. speciation, extinction and dispersal) in montane systems [6–8]. Thus, testing mechanistic hypotheses that link the high species richness of tropical montane systems to evolutionary and biogeographic processes is an important challenge for macroecologists and biogeographers.

In a highly influential paper, Janzen [9] noted that tropical mountains generally experience less seasonal variation in temperature relative to temperate mountains. This, he argued, would result in greater thermal stratification along elevational gradients in tropical montane systems than in temperate montane systems. Janzen went on to suggest that such extreme stratification selects for organisms with narrower thermal tolerances in tropical mountains, resulting in reduced dispersal over high- (and low-) elevation passes. Since the publication of his seminal paper, several studies have confirmed Janzen's predictions; in tropical montane regions species exhibit greater turnover along elevational climatic gradients, occupy more restricted elevational ranges and thermal regimes, and have narrower thermal tolerances [10–16].

Although Janzen [9] did not explicitly attempt to explain why the tropics harbour so many more species relative to other regions, there are obvious connections between his conjectures and evolutionary mechanisms promoting geographical variation in species richness [12,17]. If species from temperate-zone mountains generally inhabit broader thermal regimes (and have correspondingly broader thermal tolerances) relative to tropical species, then even the climatic differences between many mountaintops and valleys in the temperate zone may not be dramatic enough to act as dispersal barriers that promote isolation and speciation. In contrast, if tropical species adapt to narrower thermal regimes in response to the limited overlap in temperature at different elevations, then at least two mechanisms might promote greater opportunities for speciation along tropical mountains. If the thermal regimes and thermal tolerances of tropical species are more evolutionarily conserved than those of temperate species, then populations inhabiting adjacent mountaintops or valleys should experience greater isolation and increased opportunities for allopatric speciation [18]. Alternatively, the greater climatic stratification of tropical mountains might increase the likelihood of *in situ* divergence of populations and parapatric speciation along elevational climatic gradients [17,18]. The former scenario implies niche conservatism at speciation whereas the latter implies niche divergence.

These alternative pathways to tropical speciation predict contrasting patterns of evolution in the elevational distributions and thermal regimes of tropical versus temperate sister species. If thermal niches tend to be more conserved in the tropics than in the temperate zone (thereby promoting greater opportunities for allopatric speciation), then tropical sister species should inhabit similar elevational zones, and thus display a greater overlap in their thermal regimes, than temperate sister species [17]. In contrast, if the stronger elevational zonation of climate in the tropics drives speciation by promoting increased specialization of populations to non-overlapping thermal conditions, then sister species should exhibit a lesser overlap in their elevational distributions and thermal regimes in the tropics relative to the temperate zone [17].

Despite the potentially important linkages between climatic zonation and the build-up of species richness, few studies have examined the relationship between latitude, thermal-niche variation and speciation. Kozak & Wiens [17] compared the elevational and thermal overlaps of temperate and tropical sister species of plethodontid salamanders and found evidence for greater divergence in the thermal regimes of tropical species. In addition, rates of climatic niche divergence in these salamanders are positively correlated with rates of clade diversification and with latitudinal variation in species richness [19]. However, studies on other taxa have found no consistent relationship between latitude and the divergence or conservatism of sister species' thermal regimes [20], nor that rates of thermal regime divergence are actually slower in the tropics [21]. Thus, whether a general relationship between latitude, climatic zonation and speciation exists is unclear and requires further study across a wider range of taxa.

Here, we explore the relationship between spatial variation in climate and speciation in tropical and temperate mountains across 93 pairs of sister species identified from published molecular phylogenetic analyses. The selected species represent several major groups of terrestrial vertebrates and encompass taxa with a wide range of thermoregulation strategies and dispersal abilities. We used climatic information extracted from species' locality data (i) to test the prediction that tropical species occupy narrower elevational ranges and thermal regimes across their ranges than temperate species, and (ii) to examine whether tropical sister species overlap more or less in their elevational ranges and thermal regimes than do temperate sister species.

2. MATERIAL AND METHODS

To characterize the elevational and thermal regimes of species, we constructed a database of 33 304 georeferenced localities representing known occurrence sites for 190 species from five vertebrate groups (bats, birds, frogs, lizards and snakes) occurring in the New World (electronic supplementary material, appendix A). We selected these species because they were included in published (mostly molecular) phylogenetic analyses that allowed us to identify pairs of sister species for analyses. Such pairs were identified only when 80 per cent or more of the members of the clade under study (e.g. a genus) had been included in a phylogenetic analysis. To be included in the dataset, each pair of sister species had to be restricted to either the tropics or temperate zone in continental areas of the New World. Further, at least one species in each pair was restricted to montane areas (above 1000 m), but in some pairs, especially of bats, one species also extended to the lowlands. Temperate-zone species were defined as those with distributions restricted to latitudes north of 30° N and tropical-zone species as those with distributions south of 30° N. We did not consider species from temperate South America, given the marked differences in climate between hemispheres [22,23]. The degree of overlap of the geographical distributions of species was not a criterion used to include them in our database, but in general sister species were allopatrically or parapatrically distributed (electronic supplementary material, appendix A). In total, we gathered data for 95 pairs of sister species. However, because sequence

data for species in two of the pairs were lacking (identification of sister species in these cases was based on morphology-based phylogenies), only 93 species pairs were included in our analyses of sister species (51 tropical and 42 temperate). All 190 species, however, were included in our analyses of elevational range and thermal regime width (described below).

To associate known occurrence sites with environmental data, we obtained georeferenced localities for all study species from the Global Biodiversity Information Facility portal (<http://www.gbif.org/>). Using ARCGIS v. 9.2, we then superimposed them on temperature layers at a 1 km² resolution obtained from the WorldClim database [24], and on a digital elevational model at 0.0625 km² resolution obtained from the Shuttle Radar Topography Mission (<http://www2.jpl.nasa.gov/srtm/>). Prior to conducting analyses, we inspected the data for georeferencing errors and excluded obviously misplaced localities. We also discarded localities above and below the 97.5 and 2.5 percentiles of the elevation data for each species as probable outliers. We calculated the elevational range for each species as the difference between the maximum and minimum elevations across all localities.

To characterize the thermal regime of each species, we first extracted the maximum temperature of the warmest month (Bio5) and the minimum temperature of the coldest month (Bio6) for each of its collection sites. We then calculated the width of the thermal regime of each species as the difference between its maximum observed value of Bio5 and its minimum observed value of Bio6. We calculated the degree of overlap in elevational ranges of sister species by dividing the amount of overlap by the elevational range of the species with the smaller range [17]. Therefore, our metric of elevational overlap ranges from 0 (no overlap in elevation) to 1 (complete overlap).

To estimate the degree of overlap in the thermal regimes of sister species, we first estimated the monthly temperature range experienced by each species by calculating the mean minimum and mean maximum temperature across all of its collection sites. We then calculated the overall temperature overlap for each sister species pair using the formula

$$\text{temperature overlap} = \sum_{12} \left[0.5 \left(\frac{o_i}{R_{Ai}} + \frac{o_i}{R_{Bi}} \right) \right],$$

where R_{Ai} and R_{Bi} are the ambient temperature ranges experienced by species A and B, respectively, for month i , and o_i is the overlap of R_{Ai} and R_{Bi} (in °C) for month i [17]. Because the possible range of temperatures that a species can experience differs between regions (i.e. tropics versus temperate zones), this formula standardizes the temperature overlap by the width of each species's thermal regime. Thus, this index is a measure of relative overlap ranging between 0 and 1 for each month, with the total annual temperature overlap ranging between 0 and 12.

Species can restrict the breadth of temperatures they experience by regulating activity times or selecting appropriate microhabitats. Biotic interactions might also restrict a species to a subset of the temperatures that it could otherwise tolerate. Therefore, we do not assume that thermal regime widths calculated from macroclimatic conditions are equivalent to species' actual thermal tolerances. However, our approach does assume that some relationship exists between the large-scale thermal variation across the geographical

range of a species and its thermal tolerance. Previous studies have examined the relationship between thermal regime widths estimated from macroclimate data and thermal tolerance/performance breadth, and have generally found that these variables are positively correlated [12,23,25–28].

To test whether tropical species inhabit narrower elevational ranges and narrower thermal regimes than temperate sister species, we first calculated the standardized residuals of a linear regression between the number of localities sampled per species and species' elevational and temperature ranges. This approach was used to control for any effect of sample sizes on the estimated ranges. After testing for assumptions of parametric analyses, we ran one-way analyses of variance (ANOVAs) using these standardized residuals as dependent variables and region (temperate or tropical) as a factor [17]. These analyses were conducted for all taxa pooled, as well as separately for each major taxon.

Changes in the geographical ranges of species occurring after speciation can influence inferences made about the geographical and ecological contexts in which speciation occurred [29]. Because range shifts are more likely to occur over longer periods of time and because the ages of sister species may vary with respect to latitude [30], testing for regional differences in temperature and elevational overlap requires controlling for regional variation in divergence times between species. Therefore, we used analyses of covariance (ANCOVAs) to examine differences in elevational and temperature range overlap between temperate sister species and tropical sister species, while controlling for the effect of divergence times as indexed by genetic distances. If elevational and temperature overlaps are explained by regional variation in opportunities for post-speciational range shifts, rather than by differences in the degree of climatic zonation, then one would expect a significant interaction between timing of sister taxon divergence and region to explain these measures of overlap. To set up these analyses, we obtained mitochondrial DNA sequence data for all species in the analyses from GenBank (electronic supplementary material, appendix A) and calculated uncorrected p -distances (% sequence divergence) between sister species as a surrogate for divergence times. ANCOVAs included elevational and temperature overlaps as dependent variables, region (temperate or tropical) as main factor, p -distance as a covariate to control for divergence times and number of localities as a second covariate to control for differences in sample sizes. We acknowledge that rates of molecular evolution may not be constant across all species. However, using p -distance as a covariate simply assumes that there is a positive relationship between divergence time and genetic distance, not that rates of molecular divergence are constant across all sister species pairs.

We conducted sensitivity analyses to assess the robustness of our results to alternative analytical treatments of the data, including (i) restricting analyses to species with more than 20 locality records, and conducting comparisons of elevational and thermal ranges using raw data (not residuals of the regression between these ranges and number of localities); (ii) calculating elevational overlap using the same formula used to calculate temperature overlap; and (iii) calculating temperature overlap using the formula described by Janzen [9] and not that of Kozak & Wiens [17]. Because none of these procedures led to results different from those obtained using the methods described above, we do not comment on them further, but describe them in the electronic supplementary material, appendices B–G.

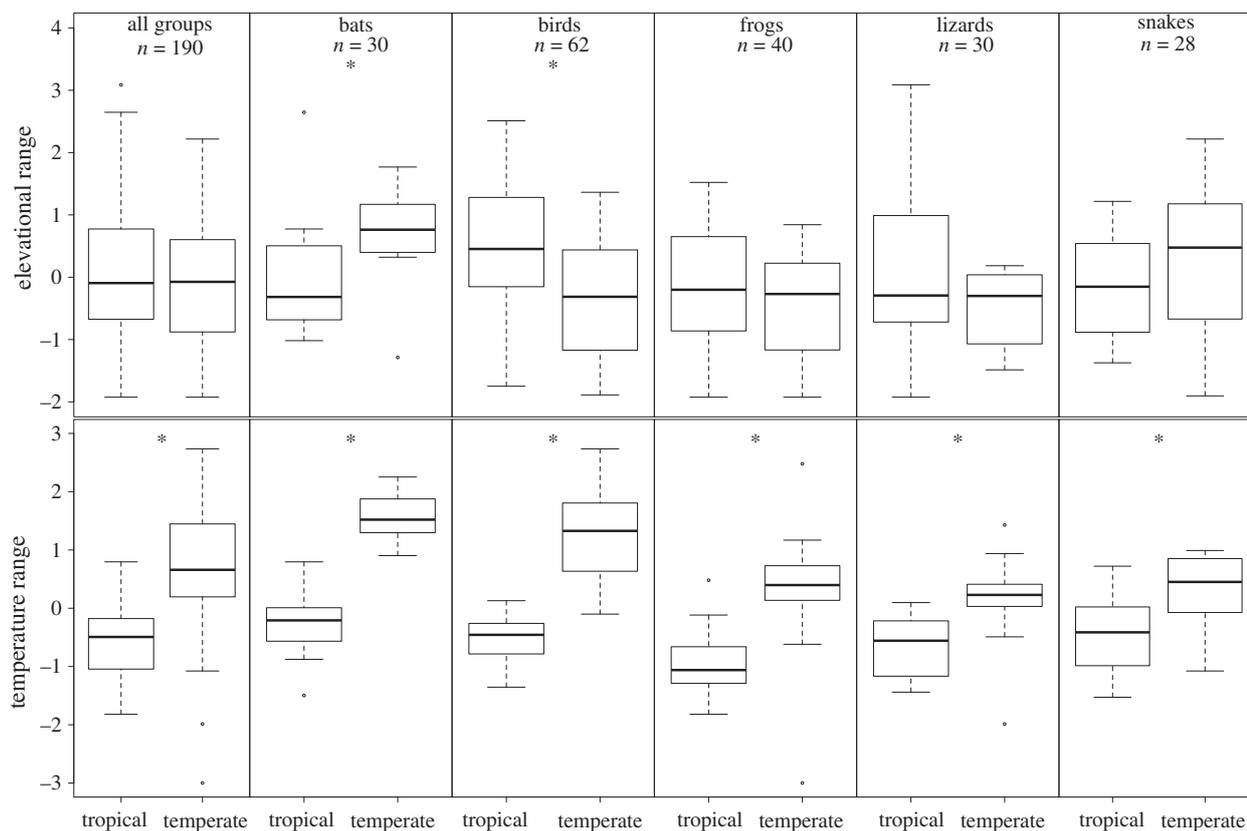


Figure 1. Elevation and temperature ranges occupied by temperate and tropical species in five vertebrate groups. Sample sizes are number of species. Boxplots show median, lower and upper quartiles, 5% and 95% percentiles, and outliers. Asterisks indicate statistically significant differences. Ranges are expressed as residuals resulting from the regressions between elevational/temperature range and number of localities sampled.

3. RESULTS

Considering all five vertebrate groups together and all species (i.e. analyses not restricted to sister species comparisons), we found no strongly significant differences in the elevational ranges of temperate versus tropical species although the analysis was marginally non-significant (ANOVA: $n = 190$, $F = 3.283$, $p = 0.072$; figure 1). When we analysed each group separately, only bats showed the expected pattern of significantly wider elevational ranges for temperate species than for tropical species ($n = 30$, $F = 3.589$, $p = 0.034$); snakes showed a similar but non-significant trend ($n = 28$, $F = 1.463$, $p = 0.237$; figure 1). The other three groups showed trends counter to the prediction, but differences in the elevational ranges of temperate and tropical species were significant only for birds (birds: $n = 60$, $F = 11.391$, $p = 0.001$; frogs: $n = 40$, $F = 0.914$, $p = 0.345$; lizards: $n = 32$, $F = 2.683$, $p = 0.112$; figure 1). By contrast, analyses combining data from all groups and independent analyses of each of the groups consistently supported the prediction that temperate species occur over significantly larger temperature ranges than do tropical species (ANOVA: all groups: $n = 190$, $F = 144.58$, $p < 0.0001$; bats: $n = 30$, $F = 109.96$, $p < 0.0001$; birds: $n = 60$, $F = 55.64$, $p < 0.0001$; frogs: $n = 40$, $F = 35.67$, $p < 0.0001$; lizards: $n = 32$, $F = 16.75$, $p < 0.0001$; snakes: $n = 28$, $F = 7.73$, $p = 0.01$; figure 1).

With respect to the overlap of sister species' elevational ranges, only temperate sister species of bats overlap less in elevational range than tropical sister species (ANCOVA: $n = 15$, $F = 3.473$, $p = 0.042$; figure 2). None of the

other analyses showed significant differences across regions in the degree of elevational overlap (ANCOVA: all groups: $n = 93$, $F = 0.575$, $p = 0.45$; birds: $n = 30$, $F = 0.726$, $p = 0.402$; frogs: $n = 20$, $F = 0.713$, $p = 0.411$; lizards: $n = 14$, $F = 0.480$, $p = 0.504$; snakes: $n = 14$, $F = 0.739$, $p = 0.410$; figure 2; full results of ANCOVAs shown in the electronic supplementary material). All groups examined showed trends indicating a greater overlap of the thermal regimes of tropical sister species relative to temperate sister species. However, results were significant only for all the groups combined, birds and lizards (ANCOVA: all groups: $n = 93$, $F = 10.42$, $p = 0.002$; lizards: $n = 14$, $F = 7.862$, $p = 0.02$; bats: $n = 15$, $F = 0.025$, $p = 0.88$; birds: $n = 30$, $F = 6.23$, $p = 0.02$; frogs: $n = 20$, $F = 0.187$, $p = 0.671$; snakes: $n = 14$, $F = 1.420$, $p = 0.26$; figure 2; full results of ANCOVAs shown in the electronic supplementary material).

4. DISCUSSION

We explored the relationship between elevational climatic zonation and speciation by comparing latitudinal patterns of variation in the thermal regimes of 93 pairs of vertebrate sister species. Consistent with the predictions of Janzen's climatic zonation hypothesis [9], we found that the geographical distributions of tropical species encompass narrower thermal regimes than those of temperate species from the Northern Hemisphere in the New World. However, in contrast to recent studies on salamanders [17] and frogs [20], we found that sister species tend to exhibit a greater overlap in their thermal

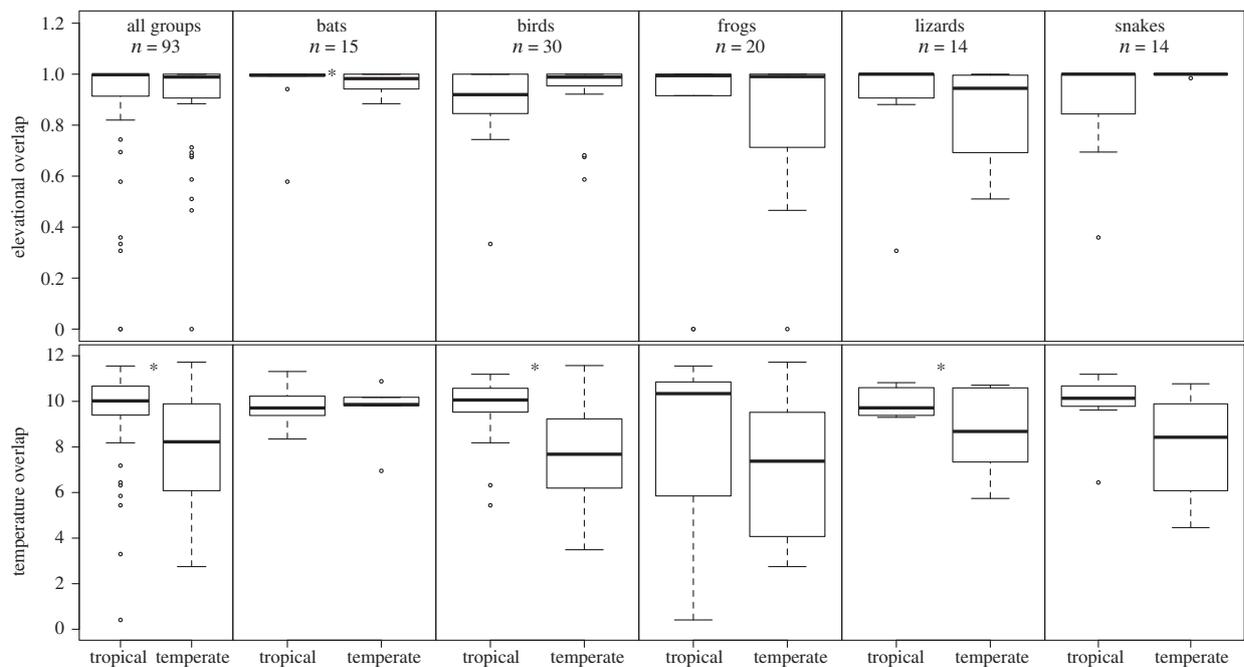


Figure 2. Overlap in elevational range and temperature regime of temperate and tropical sister species in five vertebrate groups. Sample sizes are pairs of sister species identified from molecular phylogenetic analyses. Boxplots show median, lower and upper quartiles, 5% and 95% percentiles, and outliers. Asterisks indicate statistically significant differences.

regimes in the tropics than in the temperate zone. Together, these results provide new insights on the evolutionary and biogeographic processes associated with the high species richness of tropical montane regions. Specifically, our study suggests that the thermal niches of tropical sister species tend to be narrower and more evolutionarily conserved than those of temperate sister species. As a result, populations may generally experience greater opportunities for isolation and allopatric speciation across elevational thermal gradients in tropical than in temperate montane regions.

Palaeontological and neontological studies have documented patterns of diversification consistent with the idea that rates of speciation are faster at lower latitudes (but see [30], reviewed by [31]). Some investigators have proposed that latitudinal variation in climatic zonation might contribute to this pattern by driving faster speciation in tropical montane organisms [12,17,19,20,32]. However, it has remained unclear whether the greater climatic zonation of tropical mountains might promote speciation by increasing opportunities for allopatric isolation of populations with evolutionarily conserved thermal regimes, or by driving adaptive divergence of populations distributed along elevational climatic gradients [12,17,18,20,32,33]. Based on our expanded sampling of vertebrate taxa, we find that thermal regimes of tropical sister species are generally more conserved than those of temperate ones, suggesting that the greater stability of temperature regimes along tropical mountain slopes could increase opportunities for isolation and allopatric speciation.

Of course, our results only indirectly support the idea that thermal tolerances of species are narrower and more evolutionarily conserved in tropical mountains relative to temperate ones. If species restrict the breadth of temperatures they experience by selecting appropriate microhabitats, regulating activity times or hibernating (especially at higher latitudes), then the use of

macroclimatic data could overestimate the actual thermal tolerance breadths of species. However, the few studies that have systematically examined latitudinal variation in thermal tolerance have found a positive relationship between latitude, thermal regime breadth and thermal tolerance range [13,17,23,25–28,34], although interspecific variation in thermal tolerance breadth increases with latitude as a result of the presence of species engaging in periods of extended inactivity [23]. Thus, based on available evidence, our results do not appear to be driven by a greater mismatch between thermal regime breadths and thermal tolerance ranges in the temperate zone. Regardless, explicit tests of whether thermal regime, thermal tolerance, the thermal sensitivity of performance and the ability for thermal acclimation are correlated at geographical scales are sorely needed [35].

Similarly, if biotic interactions (e.g. competition) limit the ranges of species [36,37], then thermal regime widths inferred from geographical distribution data may encompass only a subset of the thermal conditions that species can tolerate. Although biotic interactions are generally thought to be stronger in the tropics [38], there is some evidence to the contrary. For example, Huey [10] found that the turnover of species along tropical versus temperate elevational gradients is unrelated to variation in the number of co-occurring species, a finding that contradicts the idea that competition plays a greater role in driving faunal turnover in the tropics. Likewise, in treefrogs, competition appears to play a role in community assembly in the temperate zone, but not in the tropics [39]. It also seems unlikely that the latitudinal trends we document here are explained entirely by mismatches between the realized and fundamental thermal niches of species. First, given the stronger thermal gradients in tropical mountains, one would expect sister species to have divergent thermal niches, which is the exact opposite of the trend that we recovered. Second, interactions between

species should reinforce any impacts of elevational climatic zonation on the physiological tolerances of species. For example, competitive interactions would be more likely to prevent a tropical than a temperate species from encountering a wide range of climatic conditions. Over evolutionary time, one might reasonably expect such pre-emptive occupation of geographical space to cause tropical species to become more physiologically specialized than temperate species. Thus, latitudinal patterns in thermal niche evolution may ultimately arise as a result of interactions between abiotic and biotic factors [19].

In contrast to the predictions of Janzen's climatic zonation hypothesis and the results of a recent study that quantified vertebrate elevational range sizes across latitude [15], we did not find that tropical species had narrower elevational extents than temperate species. However, we point out that patterns of thermal variation, rather than elevation *per se*, formed the core of Janzen's idea. If species shift their elevational ranges locally to track preferred temperatures (e.g. moving to higher elevations at lower latitudes and *vice versa*), then range-wide measures of elevational ranges (as we report here) are likely to provide a misleading proxy for the range of thermal conditions over which a species occurs [40]. Previous studies focused on local transects or single mountains [9,10,15] have not encountered this additional complexity, which may emerge when the ranges of elevations occupied by multiple populations of any given species are evaluated. Future studies should address whether populations of elevationally wide-ranging species show a greater propensity for local thermal adaptation in the tropics relative to the temperate zone (e.g. low seasonality in the tropics may enable local adaptation of physiology at small scales more readily).

Our inferences assume that the geographical and ecological contexts of speciation have not been obscured by regional differences (i.e. tropics versus temperate zones) in opportunities for post-speciation changes in the distributions of species. By including genetic distance between species as a covariate in analyses (a surrogate for the time available for post-speciation range shifts), we have attempted to reduce this potential source of error. Genetic distances did not vary significantly between temperate and tropical species in our dataset, suggesting that regional differences in the time available for species ranges to change in position and size [30,41] do not explain the latitudinal trends that we found, although we note that the rate at which species distributions shift following speciation can vary with latitude [42,43]. Further, we cannot rule out the possibility that species in both regions have shifted their elevational and climatic distributions since their formation in response to climate change, mountain uplift or species interactions [36,44,45]. Nevertheless, it is difficult to envision how extensive post-speciation range shifts would result in a statistically significant, rather than random, pattern of the thermal overlap of tropical versus temperate sister species. Regardless, genetic studies of inter-population migration rates will be critical to further test the hypothesis that climatic gradients on mountains are stronger barriers to dispersal in the tropics compared with temperate systems.

Our results suggest that the thermal niches of tropical vertebrates are generally more conserved at the level of sister species than are those of temperate ones, a result

consistent with a recent analysis focused on mammals using a different comparative approach [21]. However, exactly why tropical species should exhibit greater conservatism of their thermal niches is unclear and will require further study. One possible explanation is that greater climatic stability in the tropics over time [46] has promoted phylogenetic conservatism in the thermal niches of species. Alternatively, but non-exclusively, biotic interactions in the tropics could restrict the climatic distributions of lineages over time [47].

Although our results suggest that climatic niches of vertebrate sister species are generally more conserved in the tropics than in the temperate zone, this pattern is not universal. For example, the narrower thermal tolerances of tropical plethodontid salamanders lead to a greater tendency for climatic niche divergence and speciation along elevational climatic gradients [17]. In general, the greater climatic zonation of tropical mountains should increase opportunities for either allopatric or parapatric speciations. However, the extent to which such climatic zonation triggers speciation along elevational gradients may ultimately depend on the balance between dispersal and selection [48]. Thus, one might predict that ecological speciation through climatic niche divergence along mountain slopes would be more prevalent in tropical taxa with the most limited dispersal abilities. Among the vertebrate taxa examined to date, plethodontid salamanders exhibit the most extreme spatial genetic structuring of populations [49], suggesting that they are probably more dispersal-limited than birds, mammals, reptiles and anurans. Thus, differences in dispersal abilities might explain the different speciation patterns between plethodontids and other vertebrate taxa.

Taken together, and bearing in mind the potential shortcomings of exploring thermal regime variation from distributional data, our results suggest that tropical sister species exhibit greater evolutionary conservatism in their thermal niches than temperate sister species. Although numerous studies have documented dramatic differences in species richness and rates of diversification between the tropics and temperate zones, few have quantified, to the extent demonstrated here, how the interplay between climatic conditions, the evolution of species' niches and speciation might shape patterns of diversity. This work, we hope, will inspire more detailed examination of the physiological mechanisms that might underlie the patterns we document here, and whether and how they influence variation in speciation mechanisms among taxa and latitudes.

This study was conducted as part of the working group Montane Biodiversity in Space and Time: Linking Evolutionary Biology and Macroecology, supported by the National Evolutionary Synthesis Center (NESCent), NSF EF-0423461. We thank the editors, CDC's laboratory group and two anonymous reviewers for helpful comments on the manuscript.

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Appendix A. Sister species of different vertebrate groups used to examine patterns of elevational distribution and climatic niche divergence in tropical and temperate montane systems of the New World. For each pair, the total number of localities used for analyses, the mtDNA gene used to calculate genetic distance (a surrogate for divergence time) and the degree of geographic overlap are indicated, together with references of phylogenetic studies from which mtDNA sequences were obtained. Additionally, the number of localities for each species is shown in parentheses.

Taxon	Distribution	Species Pair	Total No. of Localities	mtDNA Sequence	Geographic Overlap	Reference
Bats	Tropical	<i>Artibeus aztecus</i> (22) - <i>Artibeus watsoni</i> (26)	48	CytB	Allopatric	Redondo et al. 2008
		<i>Artibeus fraterculus</i> (48) - <i>Artibeus hirsutus</i> (51)	99	CytB	Allopatric	Redondo et al. 2008
		<i>Carollia brevicauda</i> (558) - <i>Carollia perspicillata</i> (691)	1249	CytB	Parapatric	Wright et al. 1999
		<i>Chrotopterus auritus</i> (45) - <i>Vampyrum spectrum</i> (21)	66	COI	Parapatric	Jones et al. 2002
		<i>Noctilio albiventris</i> (55) - <i>Noctilio leporinus</i> (75)	130	CytB	Parapatric	Lewis-Oritt et al 2001
		<i>Peropteryx kappleri</i> (18) - <i>Peropteryx macrotis</i> (55)	73	CytB	Parapatric	Lim et al. 2008
		<i>Phyllostomus elongatus</i> (83) - <i>Phyllostomus hastatus</i> (115)	198	COI	Parapatric	Jones et al. 2002
	Temperate	<i>Rhogeessa alleni</i> (13) - <i>Rhogeessa gracilis</i> (4)	17	COI	Parapatric	Baird et al. 2008
		<i>Saccopteryx bilineata</i> (335) - <i>Saccopteryx leptura</i> (59)	394	COI	Parapatric	Lim et al. 2008
		<i>Vampyressa melissa</i> (8) - <i>Vampyressa pusilla</i> (37)	45	COI	Allopatric	Porter & Baker 2004
		<i>Euderma maculatum</i> (30) - <i>Idionycteris phyllotis</i> (17)	47	ND2	Parapatric	Jones et al. 2002
		<i>Myotis californicus</i> (203) - <i>Myotis leibii</i> (154)	357	CytB	Allopatric	Stadelmann et al. 2007
		<i>Myotis evotis</i> (126) - <i>Myotis keenii</i> (75)	201	CytB	Parapatric	Stadelmann et al. 2007
		<i>Myotis lucifugus</i> (256) - <i>Myotis occultus</i> (14)	270	CytB	Allopatric	Stadelmann et al. 2007
<i>Myotis sodalis</i> (70) - <i>Myotis volans</i> (62)	132	CytB	Allopatric	Stadelmann et al. 2007		
Birds	Tropical	<i>Cinclodes aricomae</i> (2) - <i>Cinclodes excelsior</i> (17)	19	COII	Allopatric	Chesser 2004
		<i>Coeligena bonapartei</i> (31) - <i>Coeligena helianthea</i> (55)	86	ND2	Parapatric	Parra 2010
		<i>Cyanolyca turcosa</i> (43) - <i>Cyanolyca viridicyanus</i> (42)	85	ND2	Allopatric	Bonaccorso 2009
		<i>Hemispingus frontalis</i> (31) - <i>Hemispingus melanotis</i> (40)	71	CytB	Parapatric	Garcia-Moreno et al. 2001
		<i>Hemispingus verticalis</i> (13) - <i>Hemispingus xanthophthalmus</i> (15)	28	ND2	Allopatric	Garcia-Moreno et al. 2001
		<i>Henicorhina leucoptera</i> (11) - <i>Henicorhina leucosticta</i> (373)	384	ATPase 6	Allopatric	Dingle et al. 2006
		<i>Myadestes coloratus</i> (141) - <i>Myadestes ralloides</i> (105)	246	CytB	Allopatric	Miller et al. 2007
		<i>Myioborus melanocephalus</i> (115) - <i>Myioborus ornatus</i> (7)	122	CytB	Allopatric	Pérez-Emán 2005
<i>Pionus seniloides</i> (30) - <i>Pionus tumultuosus</i> (48)	78	CytB	Allopatric	Ribas et al. 2007		

Taxon	Distribution	Species Pair	Total No. of Localities	mtDNA Sequence	Geographic Overlap	Reference
		<i>Scytalopus atratus</i> (21) - <i>Scytalopus bolivianus</i> (9)	30	CytB	Allopatric	Cadena et al. Unpubl. Data
		<i>Scytalopus parkeri</i> (6) - <i>Scytalopus spillmanni</i> (23)	29	ND2	Parapatric	Cadena et al. Unpubl. Data
		<i>Tangara florida</i> (52) - <i>Tangara icterocephala</i> (269)	321	CytB	Parapatric	Burns & Naoki 2004
		<i>Tangara parzudakii</i> (78) - <i>Tangara schrankii</i> (80)	158	CytB	Allopatric	Burns & Naoki 2004
		<i>Troglodytes ochraceus</i> (112) - <i>Troglodytes solstitialis</i> (108)	220	ND2	Allopatric	Rice et al. 1999
		<i>Trogon collaris</i> (327) - <i>Trogon personatus</i> (98)	425	CytB	Allopatric	Dacosta & Klicka 2008
		<i>Veniliornis callonotus</i> (62) - <i>Veniliornis dignus</i> (14)	76	CytB	Allopatric	Moore et al. 2006
		<i>Baeolophus atricristatus</i> (11) - <i>Baeolophus bicolor</i> (79)	90	COI	Allopatric	Avise & Zink 1988
		<i>Catharus bicknelli</i> (351) - <i>Catharus minimus</i> (199)	550	CytB	Parapatric	Johnson & Cicero 2004
		<i>Cyanocitta cristata</i> (320) - <i>Cyanocitta stelleri</i> (327)	647	CytB	Allopatric	Klicka & Zink 1997
		<i>Dendroica castanea</i> (453) - <i>Dendroica fusca</i> (424)	877	CytB	Parapatric	Rabosky & Lovette 2008
		<i>Dendroica pensylvanica</i> (507) - <i>Dendroica striata</i> (629)	1136	CytB	Parapatric	Rabosky & Lovette 2008
		<i>Empidonax alnorum</i> (409) - <i>Empidonax traillii</i> (1249)	1658	CytB	Allopatric	Johnson & Cicero 2002
	Temperate	<i>Empidonax difficilis</i> (116) - <i>Empidonax occidentalis</i> (31)	147	CytB	Allopatric	Johnson & Cicero 2002
		<i>Oporornis philadelphia</i> (232) - <i>Oporornis tolmiei</i> (193)	425	CytB	Allopatric	Weir & Schluter 2004
		<i>Picoides albolarvatus</i> (22) - <i>Picoides stricklandi</i> (46)	68	CytB	Allopatric	Weibel & Moore 2002
		<i>Toxostoma bendirei</i> (57) - <i>Toxostoma cinereum</i> (67)	124	CytB	Parapatric	Zink et al. 1999
		<i>Toxostoma curvirostrae</i> (934) - <i>Toxostoma rufum</i> (255)	1189	CytB	Allopatric	Zink et al. 1999
		<i>Vermivora chrysoptera</i> (715) - <i>Vermivora pinus</i> (130)	845	CytB	Parapatric	Klein et al. 2004
		<i>Vermivora luciae</i> (122) - <i>Vermivora virginiae</i> (137)	259	CytB	Parapatric	Klein et al. 2004
		<i>Zonotrichia atricapilla</i> (432) - <i>Zonotrichia leucophrys</i> (758)	1190	CytB	Parapatric	Wickenstein et al. 2001
Frogs	Tropical	<i>Atelopus varius</i> (34) - <i>Atelopus zeteki</i> (8)	42	CytB	Parapatric	Richards & Knowles 2007
		<i>Bufo glaberrimus</i> (17) - <i>Bufo guttatus</i> (8)	25	12S	Allopatric	Pramuk 2006
		<i>Colostethus pulchellus</i> (13) - <i>Colostethus vertebralis</i> (11)	24	12S	Allopatric	Graham et al. 2004
		<i>Craugastor chac</i> (10) - <i>Craugastor lineatus</i> (4)	14	COI	Allopatric	Crawford & Smith 2005
		<i>Gastrotheca marsupiata</i> (20) - <i>Gastrotheca pseustes</i> (12)	32	12S	Allopatric	Wiens et al. 2006
		<i>Isthmohyla pseudopuma</i> (19) - <i>Isthmohyla zeteki</i> (11)	30	12S	Parapatric	Wiens et al. 2006
		<i>Isthmohyla rivularis</i> (36) - <i>Isthmohyla tica</i> (22)	58	12S	Allopatric	Wiens et al. 2006
		<i>Plectrohyla guatemalensis</i> (24) - <i>Plectrohyla matudai</i> (11)	35	CytB	Allopatric	Wiens et al. 2006
		<i>Pristimantis elegans</i> (2) - <i>Pristimantis nervicus</i> (1)	3	COI	Allopatric	Pinto et al. Unpubl. Data

Taxon	Distribution	Species Pair	Total No. of Localities	mtDNA Sequence	Geographic Overlap	Reference
		<i>Rana vibicaria</i> (25) - <i>Rana warszewitzi</i> (50)	75	12S	Parapatric	Hillis & Wilcox 2005
		<i>Ascaphus montanus</i> (31) - <i>Ascaphus truei</i> (154)	185	CytB	Allopatric	Nielson et al. 2001
		<i>Bufo californicus</i> (25) - <i>Bufo microscaphus</i> (160)	185	COI	Allopatric	Pramuk 2006
		<i>Bufo cognatus</i> (465) - <i>Bufo debilis</i> (103)	568	CytB	Parapatric	Pramuk 2006
		<i>Bufo exsul</i> (23) - <i>Bufo boreas</i> (1339)	1362	CytB	Parapatric	Pramuk 2006
		<i>Hyla avivoca</i> (11) - <i>Hyla versicolor</i> (941)	952	CytB	Parapatric	Wiens et al. 2006
	Temperate	<i>Pseudacris brachyphona</i> (25) - <i>Pseudacris brimleyi</i> (32)	57	12S	Parapatric	Moriarty-Lemmon et al. 2007
		<i>Pseudacris cadaverina</i> (346) - <i>Pseudacris regilla</i> (2168)	2514	CytB	Parapatric	Moriarty-Lemmon et al. 2007
		<i>Pseudacris feriarum</i> (73) - <i>Pseudacris triseriata</i> (867)	940	12S	Parapatric	Moriarty-Lemmon et al. 2007
		<i>Rana aurora</i> (187) - <i>Rana cascadae</i> (119)	306	CytB	Parapatric	Hillis & Wilcox 2005
		<i>Rana boylei</i> (659) - <i>Rana luteiventris</i> (325)	984	CytB	Allopatric	Hillis & Wilcox 2005
		<i>Anolis intermedius</i> (16) - <i>Anolis laevis</i> (16)	32	COI	Parapatric	Nicholson et al. 2005
		<i>Coleonyx elegans</i> (10) - <i>Coleonyx mitratus</i> (18)	28	COI	Parapatric	Jonniaux & Kumazawa 2008
		<i>Corytophanes cristatus</i> (55) - <i>Corytophanes percarinatus</i> (6)	61	COI	Parapatric	Schulte et al. 2003
	Tropical	<i>Phrynosoma braconieri</i> (4) - <i>Phrynosoma taurus</i> (9)	13	CytB	Parapatric	Leaché & McGuire 2006
		<i>Proctoporus guentheri</i> (11) - <i>Proctoporus unsaaca</i> (3)	14	12S	Allopatric	Doan et al. 2005
		<i>Stenocercus angel</i> (2) - <i>Stenocercus guentheri</i> (31)	33	COI	Allopatric	Torres-Carvajal 2007
		<i>Stenocercus humeralis</i> (7) - <i>Stenocercus varius</i> (4)	11	COI	Allopatric	Torres-Carvajal 2007
Lizards		<i>Stenocercus ornatus</i> (7) - <i>Stenocercus percultus</i> (2)	9	COI	Allopatric	Torres-Carvajal 2007
		<i>Elgaria multicarinata</i> (1445) - <i>Elgaria panamintina</i> (15)	1460	COI	Parapatric	Macey et al. 1999
		<i>Eumeces fasciatus</i> (366) - <i>Eumeces laticeps</i> (32)	398	CytB	Parapatric	Macey et al. 2006
	Temperate	<i>Eumeces gilberti</i> (593) - <i>Eumeces skiltonianus</i> (904)	1497	COI	Parapatric	Macey et al. 2006
		<i>Gambelia sila</i> (84) - <i>Gambelia wislizeni/copei</i> (736)	820	CytB	Parapatric	McGuire et al. 2007
		<i>Phrynosoma ditmarsii</i> (6) - <i>Phrynosoma hernandesi</i> (137)	143	CytB	Parapatric	Leaché & McGuire 2006
		<i>Phrynosoma mcallii</i> (60) - <i>Phrynosoma platyrhinos</i> (789)	849	CytB	Parapatric	Hodges & Zamudio 2004
Snakes	Tropical	<i>Bothriechis aurifer</i> (8) - <i>Bothriechis nigroviridis</i> (10)	18	CytB	Parapatric	Wuster et al. 2008
		<i>Bothrocophias hyoprora</i> (15) - <i>Bothrocophias microphthalmus</i> (2)	17	CytB	Parapatric	Castoe & Parkinson 2006
		<i>Leptodeira punctata</i> (94) - <i>Leptodeira splendida</i> (11)	105	CytB	Parapatric	Mulcahy 2007

Taxon	Distribution	Species Pair	Total No. of Localities	mtDNA Sequence	Geographic Overlap	Reference
		<i>Leptomicrurus narducii</i> (27) - <i>Micrurus surinamensis</i> (15)	42	CytB	Parapatric	Castoe et al. 2007
		<i>Pituophis deppei</i> (26) - <i>Pituophis lineaticollis</i> (22)	48	COI	Parapatric	Rodriguez-Robles & De Jesus-Escobar 2000
		<i>Thamnophis chrysocephalus</i> (8) - <i>Thamnophis fulvus</i> (25)	33	CytB	Parapatric	de Queiroz et al. 2002
		<i>Thamnophis eques</i> (50) - <i>Thamnophis marcianus</i> (214)	264	CytB	Parapatric	de Queiroz et al. 2002
		<i>Thamnophis melanogaster</i> (26) - <i>Thamnophis validus</i> (75)	101	CytB	Parapatric	de Queiroz et al. 2002
		<i>Charina bottae</i> (230) - <i>Charina trivirgata</i> (163)	393	CytB	Parapatric	Slowinski & Lawson 2002
		<i>Lampropeltis getula</i> (781) - <i>Stilosoma extenuatum</i> (3)	784	COI	Parapatric	Pyron & Burbink 2009
	Temperate	<i>Lampropeltis zonata</i> (179) - <i>Lamproletis pyromelana</i> (38)	217	CytB	Allopatric	Pyron & Burbink 2009
		<i>Thamnophis atratus</i> (308) - <i>Thamnophis elegans</i> (1343)	1651	CytB	Allopatric	de Queiroz et al. 2002
		<i>Thamnophis couchii</i> (314) - <i>Thamnophis gigas</i> (31)	345	CytB	Parapatric	de Queiroz et al. 2002
		<i>Thamnophis proximus</i> (295) - <i>Thamnophis sauritus</i> (107)	402	CytB	Allopatric	de Queiroz et al. 2002

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Appendix B. Results of analyses of variance (ANOVAs) testing for differences in the (i) elevational and (ii) temperature ranges of tropical and temperate species (distribution effect) of vertebrates excluding species with less than 20 records to reduce the effect of considering species with few locality data, which can appear to occupy narrow ranges owing to statistical artifacts and not to niche restrictions. In these analyses, the effect of sample sizes was not accounted for further.

(i)

	n	F	p
Elevational Range			
All Groups			
<i>Distribution</i>	129	4.62	0.03
Bats			
<i>Distribution</i>	21	2.43	0.13
Birds			
<i>Distribution</i>	48	14.54	<0.01
Frogs			
<i>Distribution</i>	24	1.15	0.29
Lizards			
<i>Distribution</i>	14	5.35	0.04
Snakes			
<i>Distribution</i>	18	2.86	0.11

(ii)

	n	F	p
Temperature Range			
All Groups			
<i>Distribution</i>	135	13.68	<0.01
Bats			
<i>Distribution</i>	24	3.80	0.06
Birds			
<i>Distribution</i>	52	14.66	<0.01
Frogs			
<i>Distribution</i>	25	0.38	0.54
Lizards			
<i>Distribution</i>	14	0.00	0.97
Snakes			
<i>Distribution</i>	18	8.39	<0.01

Appendix C. Results of analyses of covariance (ANCOVAs) testing for differences in the (i) elevational and (ii) temperature ranges of tropical and temperate species of vertebrates (distribution effect) excluding species with less than 20 records and including the number of localities as a covariate. In these analyses, the effect of sample sizes was taken into consideration explicitly by removing species with few data and then correcting for the effect of sample sizes including this covariate (localities effect).

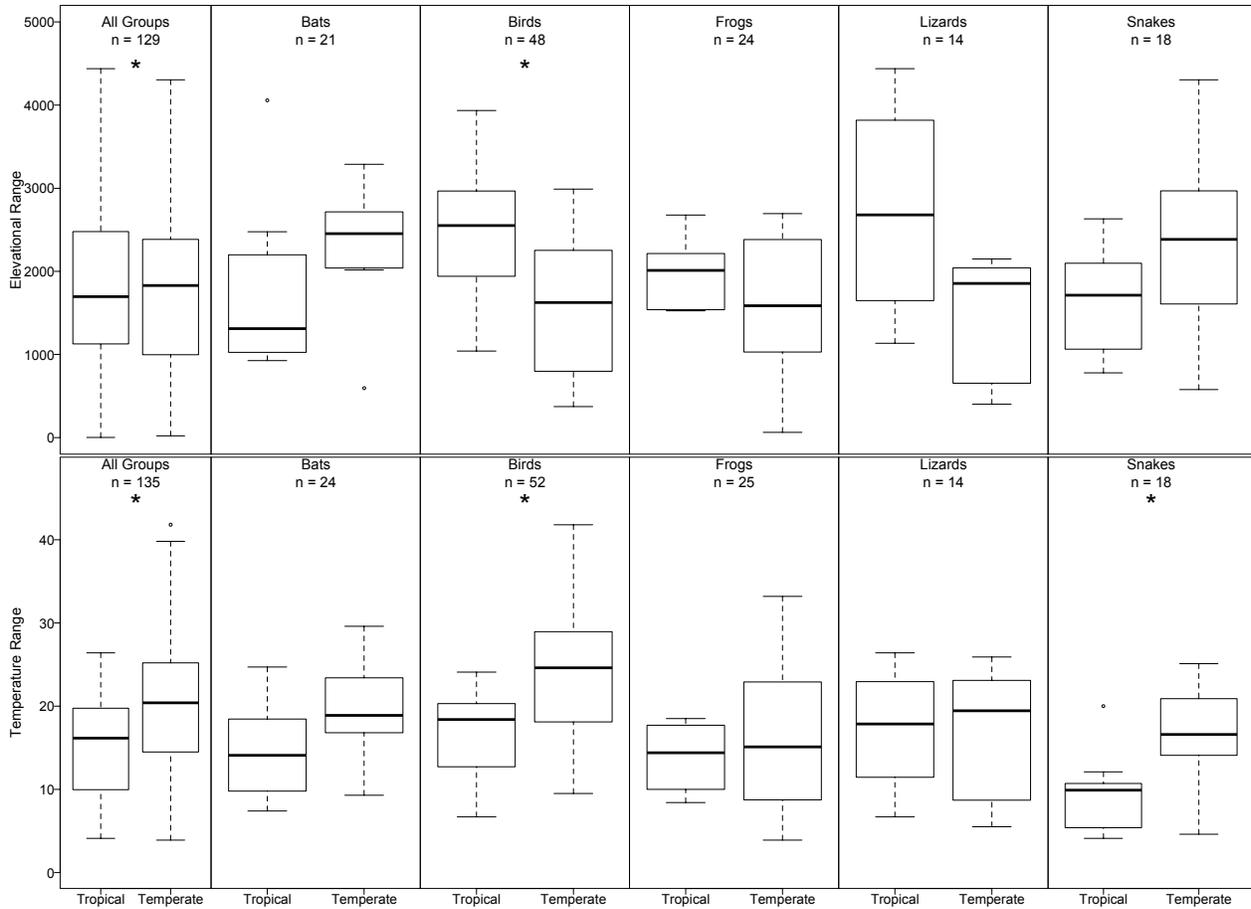
(i)

	n	F	p
Elevational Range			
All Groups			
<i>Localities</i>	129	5.02	0.02
<i>Distribution</i>	129	4.77	0.03
Bats			
<i>Localities</i>	21	0.19	0.66
<i>Distribution</i>	21	2.33	0.14
Birds			
<i>Localities</i>	48	1.03	0.31
<i>Distribution</i>	48	14.55	<0.01
Frogs			
<i>Localities</i>	24	6.35	0.01
<i>Distribution</i>	24	1.43	0.24
Lizards			
<i>Localities</i>	14	2.49	0.14
<i>Distribution</i>	14	6.02	0.03
Snakes			
<i>Localities</i>	18	17.18	<0.01
<i>Distribution</i>	18	5.43	0.03

(ii)

	n	F	p
Temperature Range			
All Groups			
<i>Localities</i>	135	31.95	<0.01
<i>Distribution</i>	135	16.86	<0.01
Bats			
<i>Localities</i>	24	4.47	0.04
<i>Distribution</i>	24	4.40	0.04
Birds			
<i>Localities</i>	52	11.27	<0.01
<i>Distribution</i>	52	17.68	<0.01
Frogs			
<i>Localities</i>	25	14.96	<0.01
<i>Distribution</i>	25	0.61	0.44
Lizards			
<i>Localities</i>	14	11.68	<0.01
<i>Distribution</i>	14	0.00	0.95
Snakes			
<i>Localities</i>	18	8.12	0.01
<i>Distribution</i>	18	11.71	<0.01

Appendix D. Elevational and temperature ranges occupied by temperate and tropical species in five vertebrate groups. Sample sizes are number of species. In contrast to the figure portrayed in the main text, which used as response variable the residuals of the regression used to correct for the number of localities, this figure shows the raw data of elevational and temperature ranges. Boxplots indicate median, lower and upper quantiles, the 5% and 95% percentiles, and outliers. Asterisks indicate significant differences based on the ANOVA shown in Appendix B. Patterns are similar to those of analyses described in the main text.



Appendix E. Full results of analyses of covariance (ANCOVAs) testing for differences in the (i) elevational and (ii) temperature overlap of sister species of vertebrates between the tropical and temperate zones (distribution effect). Results related to the tropical vs. temperate comparisons were reported in the main text, but those related to the effect of genetic distance (surrogate for divergence time) and number of localities were not reported for brevity and because they were not significant and were included in analyses only as possibly confounding variables.

(i)

	n	F	p
Elevational Overlap			
All Groups			
<i>Genetic Distance</i>	93	0.21	0.65
<i>Localities</i>	93	3.76	0.06
<i>Distribution</i>	93	0.58	0.45
Bats			
<i>Genetic Distance</i>	15	0.00	0.97
<i>Localities</i>	15	0.32	0.59
<i>Distribution</i>	15	3.47	0.04
Birds			
<i>Genetic Distance</i>	30	8.39	0.01
<i>Localities</i>	30	1.84	0.19
<i>Distribution</i>	30	0.73	0.40
Frogs			
<i>Genetic Distance</i>	20	0.23	0.64
<i>Localities</i>	20	2.27	0.15
<i>Distribution</i>	20	0.71	0.41
Lizards			
<i>Genetic Distance</i>	14	0.00	0.99
<i>Localities</i>	14	0.36	0.56
<i>Distribution</i>	14	0.48	0.50
Snakes			
<i>Genetic Distance</i>	14	0.09	0.77
<i>Localities</i>	14	0.01	0.92
<i>Distribution</i>	14	0.74	0.41

(ii)

	n	F	p
Temperature Overlap			
All Groups			
<i>Genetic Distance</i>	93	0.58	0.45
<i>Localities</i>	93	1.15	0.29
<i>Distribution</i>	93	10.42	0.00
Bats			
<i>Genetic Distance</i>	15	0.58	0.46
<i>Localities</i>	15	1.73	0.22
<i>Distribution</i>	15	0.03	0.88
Birds			
<i>Genetic Distance</i>	30	0.72	0.40
<i>Localities</i>	30	1.15	0.29
<i>Distribution</i>	30	6.23	0.02
Frogs			
<i>Genetic Distance</i>	20	0.00	0.98
<i>Localities</i>	20	0.02	0.89
<i>Distribution</i>	20	0.19	0.67
Lizards			
<i>Genetic Distance</i>	14	0.26	0.62
<i>Localities</i>	14	4.06	0.07
<i>Distribution</i>	14	7.86	0.02
Snakes			
<i>Genetic Distance</i>	14	0.16	0.70
<i>Localities</i>	14	0.08	0.79
<i>Distribution</i>	14	1.42	0.26

Appendix F. Full results of analyses of covariance (ANCOVAs) testing for differences in the (i) elevational and (ii) temperature overlap of sister species of vertebrates between the tropical and temperate zones computed using approaches different from those described in the main text. Here, elevational overlap was calculated using the formula proposed by Kozak and Wiens to calculate thermal overlap (reference 17 in the main text), whereas thermal overlap was calculated using the formula used by Janzen (9). The formulae to calculate overlap are shown in the tables, where O_{AB} is the overlap between sister species A and B, R_A is the range (in meters or degrees Celsius) of species A, and R_B is the range of species B.

(i)

	n	F	p
Elevational Overlap			
All Groups			
<i>Genetic Distance</i>	93	3.17	0.12
<i>Localities</i>	93	4.06	0.04
<i>Distribution</i>	93	0.00	0.94
Bats			
<i>Genetic Distance</i>	15	0.44	0.51
<i>Localities</i>	15	0.27	0.61
<i>Distribution</i>	15	0.01	0.91
Birds			
<i>Genetic Distance</i>	30	4.20	0.77
<i>Localities</i>	30	0.96	0.33
<i>Distribution</i>	30	0.08	0.77
Frogs			
<i>Genetic Distance</i>	20	0.35	0.55
<i>Localities</i>	20	2.15	0.16
<i>Distribution</i>	20	0.00	0.93
Lizards			
<i>Genetic Distance</i>	14	0.13	0.71
<i>Localities</i>	14	1.70	0.21
<i>Distribution</i>	14	0.03	0.86
Snakes			
<i>Genetic Distance</i>	14	0.25	0.62
<i>Localities</i>	14	0.03	0.86
<i>Distribution</i>	14	0.00	0.96

Calculated as $0.5(O_{AB}/R_A + O_{AB}/R_B)$

(ii)

	n	F	p
Temperature Overlap			
All Groups			
<i>Genetic Distance</i>	93	0.59	0.44
<i>Localities</i>	93	0.29	0.58
<i>Distribution</i>	93	9.69	0.00
Bats			
<i>Genetic Distance</i>	15	0.60	0.43
<i>Localities</i>	15	1.78	0.20
<i>Distribution</i>	15	0.12	0.73
Birds			
<i>Genetic Distance</i>	30	0.38	0.53
<i>Localities</i>	30	2.11	0.15
<i>Distribution</i>	30	6.10	0.02
Frogs			
<i>Genetic Distance</i>	20	0.00	0.98
<i>Localities</i>	20	0.02	0.87
<i>Distribution</i>	20	0.44	0.51
Lizards			
<i>Genetic Distance</i>	14	0.01	0.91
<i>Localities</i>	14	4.19	0.06
<i>Distribution</i>	14	3.90	0.07
Snakes			
<i>Genetic Distance</i>	14	0.09	0.75
<i>Localities</i>	14	0.06	0.79
<i>Distribution</i>	14	2.73	0.12

Calculated as $O_{AB}/\sqrt{(R_A * R_B)}$

Appendix G. Overlap in elevational range and temperature regime of temperate and tropical species in five vertebrate groups. Sample sizes are number of species. Here, elevational and temperature overlap were calculated as described in Appendix F (i.e. using different formulae from those used in the main text). Boxplots indicate median, lower and upper quantiles, the 5% and 95% percentiles, and outliers. Asterisks indicate significant differences based on the ANCOVA shown in Appendix F.

